BRIEF REPORT

Dolutegravir Resistance in Malawi's National HIV Treatment Program

Joep J. van Oosterhout,^{1,2} Chifundo Chipungu,¹ Lyse Nkhoma,³ Hope Kanise,¹ Mina C. Hosseinipour,⁴ Jean Babtiste Sagno,⁵ Katherine Simon,^{6,7} Carrie Cox,^{6,7} Risa Hoffman,² Kim Steegen,^{8,9} Bilaal W. Matola,¹⁰ Sam Phiri,^{1,10} Andreas Jahn,¹¹ Rose Nyirenda,¹¹ and Tom Heller³

¹Partners in Hope, Lilongwe, Malawi, ²Division of Infectious Diseases, David Geffen School of Medicine, University of California, Los Angeles, California, USA, ³The Lighthouse Trust, Lilongwe, Malawi, ⁴University of North Carolina Project Malawi, Lilongwe, Malawi, ⁵DREAM, Communion of St. Egidio, Blantyre, Malawi, ⁶Baylor College of Medicine Children's Foundation-Malawi, Lilongwe, Malawi, ⁷Baylor International Pediatric AIDS Initiative, Houston, Texas, USA, ⁸Department of Haematology & Molecular Medicine, National Health Laboratory Service, Johannesburg, South Africa, ⁹Department of Public Health and Family Medicine, Kamuzu University of Health Sciences, Lilongwe, Malawi, and ¹¹Department of HIV-AIDS, Ministry of Health, Lilongwe, Malawi,

Dolutegravir HIV drug resistance (HIVDR) data from Africa remain sparse. We reviewed HIVDR results of Malawians on dolutegravir-based antiretroviral therapy (November 2020– September 2021). Of 6462 eligible clients, 33 samples were submitted to South Africa, 27 were sequenced successfully, and 8 (30%) had dolutegravir HIVDR. Malawi urgently requires adequate HIVDR testing capacity.

Keywords. Africa; antiretroviral therapy; dolutegravir, HIV; Malawi; resistance.

Dolutegravir, a well-tolerated and highly effective antiretroviral drug, is recommended in first- and second-line antiretroviral therapy (ART) by the World Health Organization [1]. A major advantage of dolutegravir is its high genetic barrier to the development of HIV drug resistance (HIVDR) [2]. Dolutegravir resistance did not develop among ART-naïve participants in landmark trials [3, 4] and has been described in only a very few ART-naïve people with HIV (PWH) who started dolutegravirbased first-line ART [5]. However, dolutegravir resistance can develop, particularly in persons with previous exposure to older integrase inhibitors with low genetic barriers to resistance development or those with high-level resistance to the drugs used in the nucleoside backbone, resulting in dolutegravir monotherapy [6]. The risk of dolutegravir resistance may also be

Open Forum Infectious Diseases[®]2022



increased by infection with a non-B HIV subtype, high viral load (VL) and low CD4 cell count, insufficient adherence to ART, and drug interactions or malabsorption, which reduce dolutegravir drug levels [5]. These risk factors are common in Sub-Saharan Africa, but dolutegravir resistance data from the region are sparse. We therefore sought to describe dolutegravir resistance mutations in the routine setting of the Malawi HIV treatment program.

METHODS

Dolutegravir-based regimens were introduced in Malawi in 2019 [7]. Since then, a rapid transition from non-nucleoside reverse transcriptase inhibitor (NNRTI)-based (primarily efavirenz) to dolutegravir-based first-line ART has taken place. The Malawi treatment guidelines did not require documentation of viral suppression as a condition for transitioning to dolutegravir-based regimens, nor a change of the NRTI backbone if a VL result was available and elevated. Therefore, many clients may have switched while viremic and with undetected HIVDR. At the end of March 2021, >838 000 Malawians (96% of ~871 000 PWH alive on ART) in the national program were on dolutegravir-based regimens [8]. When individuals on dolutegravir-based regimens develop virological failure, defined as a second VL result of >1000 copies/mL after a period of 3 months of intensive adherence support, Malawi HIV guidelines require evidence of HIVDR before switching to a next-line ART regimen. Data from the Malawi Laboratory Information Management System from November 2020 through August 2021 indicate that 6462 samples from individuals on dolutegravirbased regimens had virological failure and were eligible for HIVDR testing. Applications for HIVDR testing need to be submitted to a national HIVDR committee [9], which determines eligibility for sample transportation to the National Health Laboratory Service, Johannesburg, South Africa. At this laboratory, RNA is extracted from dried blood spot (DBS) samples, which are stored at -80°C before testing. Two DBS samples (75 μ L each) are added to 2 mL of lysis buffer for RNA extraction using NucliSENS easyMAG. HIVDR testing is performed using previously validated in-house protocols adapted from Zhou et al. and Van Laethem et al. [10, 11]. Partial pol gene sequences are assembled and edited using RECall (British Columbia's Centre for Excellence in HIV/AIDS Research). Sequences are loaded onto the Stanford HIVdb, version 9.0, genotypic resistance system (https://hivdb.stanford.edu/hivdb/by-sequences/) to generate resistance reports.

We reviewed all cases submitted to Malawi's HIVDR expert committee and approved for integrase gene sequencing as part of HIVDR testing from the time of dolutegravir rollout,

Received 21 December 2021; editorial decision 4 March 2022; accepted 4 April 2022; published online 5 April 2022.

Correspondence: J. J. van Oosterhout, Partners in Hope, PO Box 302, Lilongwe, Malawi (joep@pihmalawi.com).

[©] The Author(s) 2022. Published by Oxford University Press on behalf of Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (https://creativecommons.org/ licenses/by-nc-nd/4.0/), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact journals.permissions@oup.com https://doi.org/10.1093/ofid/ofac148

including client characteristics, current health status, ART history, comorbidities and comedications, adherence measures, VL results, and genotyping results. In Malawi, integrase gene sequencing is only allowed for clients with current exposure to integrase inhibitors.

Patient Consent

The National Health Science Research Committee, Lilongwe, Malawi, waived individual informed consent and provided ethical approval for analysis and dissemination of the routinely collected, anonymized data presented.

RESULTS

Eighty-seven applications for HIVDR testing of clients on dolutegravir were received between November 12, 2020, and September 2, 2021. Of these, 34 were not accepted due to ineligibility (50% because of indications of current poor adherence), and 20 samples could not be transported to South Africa due to coronavirus disease 2019 (COVID-19)-related suspension of air flights. Samples from 33 PWH underwent resistance testing including integrase gene sequencing. Of these, 4 clients were on treatment at central hospitals, 21 at rural, mission, or district hospitals, and 8 at health centers. All clients had HIV subtype C. We identified dolutegravir resistance mutations from 8 of the 27 samples that were successfully sequenced (6 did not amplify). Clinical details and HIVDR testing results of the 8 cases are summarized in Tables 1 and 2. Seven were male (88%); the ages ranged from 15 to 46 years, and all were on a single-tablet generic combination of dolutegravir/tenofovir disoproxil fumarate/lamivudine. At initiation of that regimen, 1 patient was ART naïve, 4 had switched from non-dolutegravir-based firstline ART, and 3 from second-line ART. The median duration of viremia on dolutegravir was 12.5 months. Recommendations for next-line regimens were provided based on HIVDR test results and included tenofovir/lamivudine plus a double dose of dolutegravir and/or darunavir/ritonavir.

DISCUSSION

Our cases demonstrate the emergence of dolutegravir resistance among Malawians who are treated in routine settings of the national HIV treatment program. In a 2019 observational study with >1300 Malawians who had transitioned to a dolutegravir-based regimen and were assessed 6 months later, 98% achieved VL suppression. Of 6 clients with confirmed virological failure, 2 had dolutegravir resistance. These results documented early dolutegravir resistance development among clients who had transitioned to dolutegravir-based regimens from mainly NNRTI-based regimens [13]. While our finding of 8 cases with dolutegravir resistance mutations among 27 PWH (30%) with integrase sequencing results may suggest that dolutegravir resistance is common in the Malawi HIV program among individuals with virological failure, the true prevalence is currently unknown.

Of 6462 clients who were eligible for integrase sequencing from November 2020 through August 2021, only 33 samples were actually sent for HIVDR testing, indicating the practical and logistical challenges of HIVDR testing procedures, such as limited awareness of HIVDR testing indications and procedures, lack of local integrase sequencing capacity, insufficient funding, and air flight restrictions for sample transport during the COVID-19 pandemic. These restrictions may have limited switching to second-line treatment.

Several factors may increase the development of dolutegravir drug resistance in the Malawi HIV program. First, many individuals in Malawi who transitioned from NNRTI- to dolutegravir-based regimens did not have a VL result at the time of switching (as observed in 5 of our 8 cases) and continued the same NRTI backbone, creating the potential for functional dolutegravir monotherapy. Surprisingly, we did not observe intermediate/high-level resistance to tenofovir in most of our cases (Table 2). The NADIA study showed that in similar settings as ours, high VL suppression was achieved with dolutegravir regimens containing NRTIs with no activity as predicted by genotyping. Of concern and in line with our findings, 4 of 14 participants with viral rebound among 441 NADIA participants in the dolutegravir arm developed intermediate- or high-level dolutegravir resistance [14]. Second, considerable delays in the management of virological failure and HIVDR testing procedures were observed among our cases, leading to long-term viremia during dolutegravir exposure, which may facilitate progressive accumulation of dolutegravir resistance mutations. Five of our cases had detectable viremia for more than a year before HIVDR testing was done. These findings underline that enhanced VL testing is needed to improve virological failure management and prevent HIVDR development. Lastly, all samples that amplified, with and without dolutegravir resistance, demonstrated HIV subtype C, which is the predominant HIV strain among Malawian PWH [15], and non-B subtypes may increase the risk of dolutegravir resistance [5].

Client 1 was reportedly ART naïve and developed dolutegravir resistance on first-line dolutegravir/tenofovir disoproxil fumarate/lamivudine, which according to the literature is extremely rare [5]. The presence of NNRTI drug resistance mutations suggests transmitted NNRTI resistance or nondisclosure of previous exposure to ART. Investigators from the ADVANCE study in South Africa have proposed that virological failure on a dolutegravir-based regimen may be facilitated by baseline NNRTI resistance through an unknown mechanism [16].

We did not observe protease inhibitor (PI) resistance in any of the samples of the 8 cases. Five individuals had never been exposed to PIs, while the 3 who had been on PI-based secondline ART before switching to a dolutegravir-based regimen may

NAOptimalOptimalGood4No VL availableSuboptimalOptimalGood13No VL availableOptimalGoodGood13No VL availableOptimalGoodGood28No VL availableSuboptimalOptimalOptimal28No VL availableOptimalOptimalGood28No VL availableOptimalOptimalGood28No VL availableOptimalOptimalGood28No VL availableSuboptimalNo info28Sa00 000SuboptimalSuboptimalNo info12Sa00 000SuboptimalSuboptimalNo info12Sa00 000SuboptimalSuboptimalSuboptimal8	Client Number	Age ^a	Sex	Total Duration ART, mo	Current Regimen (Duration, mo)	On First- or Second-Line Regimen ^b	Previous ART Regimens ^c	Virological Status at Switch to DTG Regimen	Pill Count Adherence ^d	Self- Reported Adherence ^e	Adherence As- sessment After IAC Sessions ^f	Viremia Dura- tion on DTG Regimen, mo ^g	Clinical Status	Potential DTG Drug Interactions
30 M 123 TPF3TC/ DTG (23) 1st dTJ3TC/NVF TDFJ3TC/FVV No Lavailable Suboptimal Good 13 38 M 12 TDF3TC/ DTG (23) 1st dTJ3TC/NVF TDFJ3TC/FVV No Lavailable Suboptimal Good 13 20 M 125 TDF3TC/ DTG (23) 1st dTJ3TC/NVF No Lavailable Suboptimal Good 13 41 M 30 TDF3TC/ DTG (23) 1st dTJ3TC/NVF No Lavailable Suboptimal Good 13 41 M 30 TDF3TC/ DTG (24) 1st UL available Suboptimal Good 13 41 M 30 TDF3TC/ DTG (24) No L available Optimal Optimal Good 20 41 M 19 TDF3TC/ DTG (24) No L available Optimal Optimal Good 20 41 M 19 UTG (24) No L available Optimal Optimal Good 20 41 M 19 UTG (24) No L available Optimal Optimal Good <td>-</td> <td>46</td> <td>Σ</td> <td>29</td> <td>TDF/3TC/ DTG (29)</td> <td>1st</td> <td>None</td> <td>AA</td> <td>Optimal</td> <td>Optimal</td> <td>Good</td> <td>4</td> <td>CD4 = 61; HBsAg positive, weight loss</td> <td>Unknown herbal medications</td>	-	46	Σ	29	TDF/3TC/ DTG (29)	1st	None	AA	Optimal	Optimal	Good	4	CD4 = 61; HBsAg positive, weight loss	Unknown herbal medications
38 M 121 DF3TC(DTG (23) 1st d473TC/NVF, DTG (23) NoVL available Optimal Good 03 20 M 125 DF/3TC(3) 1st d473TC/NVF, AZT/3TC/NVF NoVL available Suboptimal Optimal Good 28 41 M 30 DTF/3TC(3) 1st UTF/3TC/NVF NoVL available Optimal Optimal 0ptimal 0ptimal 28 41 M 30 DTF/3TC/ 1st UTF/3TC/FFV NoVL available Optimal Optimal 0ptimal 0ptimal 28 42 M 179 DTF/3TC/FFV NoVL available Optimal Optimal 0ptimal 0ptimal 30 30 15 F 26 DTF/3TC AT/3TC/FFV NoVL available Suboptimal Optimal 30 30 16 N 176 Suboptimal Optimal Optimal Optimal 0ptimal 30 30 17 176 NoVL available Suboptimal Optimal Optimal 30 30 30 30	2	39	Σ	123	TDF/3TC/ DTG (23)	1st	d4T/3TC/NVP, TDF/3TC/EFV	No VL available	Suboptimal	Optimal	Good	13	No CD4 count available; asymptomatic	None
20 M 125 TDF/3TC/NUE 1st d47/3TC/NUE No VL available Suboptimal No info 28 41 M 30 TDF/3TC/ 1st TDF/3TC/NUE No VL available Optimal No info 28 42 M 179 TDF/3TC/ 2nd d47/3TC/NUE No VL available Optimal Good 20 42 M 179 TDF/3TC/ 2nd d47/3TC/NUE No VL available Suboptimal Good 20 41 TDF/3TC/EFV No VL available Suboptimal Optimal Good 20 41 TDF 2nd d47/3TC/FFV No VL available Suboptimal Good 20 41 TDF/3TC/FFV 2nd d47/3TC/FFV No VL available Suboptimal Suboptimal 30 45 M 126 DTG/3TC/FFV 2nd d47/3TC/FFV Suboptimal Suboptimal 30 46 M 126 DTG/3TC/FFV Suboptimal Suboptimal Suboptimal 30 47 DTG/3D 2nd	e	38	Σ	121	TDF/3TC/ DTG (23)	1st	d4T/3TC/NVP, TDF/3TC/EFV	No VL available	Optimal	Good	Good	13	No CD4 count available; asymptomatic	None
41 M 30 TDF/3TC/ DTG (24) 1st TDF/3TC/ TDF/3TC/FV No VL available Optimal Good 20 42 M 179 TDF/3TC/ DTG (7) 2nd d47/3TC/FV No VL available Suboptimal Good 3 15 F 26 TDF/3TC/ DTG (9) 2nd AZT/3TC + LPV/r >300 000 Suboptimal Suboptimal No info 3 46 M 126 TDF/3TC/FV >300 000 Suboptimal Suboptimal No info 12 46 M 126 TDF/3TC/FV >22 000 copies/mL Suboptimal Suboptimal Suboptimal Suboptimal 8 12	4	20	Σ	125	TDF/3TC/ DTG (28)	1st	d4T/3TC/NVP, AZT/3TC/NVP	No VL available	Suboptimal	Optimal	No info	28	No CD4 count available; asymptomatic	None
42 M 179 TDF/3TC/ 2nd d4T/3TC/NVP, No VL available Suboptimal Good 3 15 F 26 TDF/3TC/ 2nd ABC/3TC + ATV/r >300 000 Suboptimal No info 12 46 M 126 TDF/3TC/ 2nd 47/3TC/FFV >200 000 Suboptimal No info 12 46 M 126 TDF/3TC/FFV >200 copies/mL No No 12 12 46 M 126 TDF/3TC/FFV >22 000 copies/mL Suboptimal Suboptimal Suboptimal 80 potimal 8	ŋ	41	Σ	00	TDF/3TC/ DTG (24)	1st	TDF/3TC/EFV	No VL available	Optimal	Optimal	Good	20	No CD4 count available; asymptomatic	None
15 F 26 TDF/3TC/ 2nd ABC/3TC + LPV/r >300 000 Suboptimal No info 12 Nc 46 M 126 TDF/3TC/ 2nd d4T/3TC/NVP, >>22 000 copies/ Suboptimal Suboptimal No info 12 Nc 46 M 126 TDF/3TC/ 2nd d4T/3TC/NVP, >>22 000 copies/ Suboptimal Suboptimal Suboptimal 8 CC	9	42	Σ	179	TDF/3TC/ DTG (7)	2nd	d4T/3TC/NVP, TDF/3TC/EFV, AZT/3TC + ATV/r	No VL available	Suboptimal	Optimal	Good	ო	CD4 = 70; weight loss; active EPTB	Rifampicin; DTG dose was doubled during TB treatment
46 M 126 TDF/3TC/ 2nd d4T/3TC/NVP, >22 000 copies/ Suboptimal Suboptimal 8 CD DTG(8) TDF/3TC/FEV mL ATT/2TC + ATV/F	2	15	ш	26	TDF/3TC/ DTG (9)	2nd	ABC/3TC + LPV/r	>300 000 copies/mL	Suboptimal	Suboptimal	No info	12	No CD4 count available; asymptomatic	None
	œ	46	Σ	126	TDF/3TC/ DTG (8)	2nd	d4T/3TC/NVP, TDF/3TC/EFV AZT/3TC + ATV/r	>22 000 copies/ mL	Suboptimal	Suboptimal	Suboptimal	ω	CD4 = 180; asymptomatic	None

Table 1. Clinical Details of Patients With DTG Resistance Mutations

^aAge is at the time of application for genotyping.

^bSecond line: switched from PLbased second-line regimen to TDF/3TC/DTG, as recommended in national guidelines.

cincluding single-drug changes due to side effects and listed from oldest to most recent.

^dOptimal = 95%-105% pill count adherence; suboptimal = any value outside the optimal range. *Optimal = positive response to all 3 questions; suboptimal = negative response to any of 3 questions.

^fNarrative description by submitting clinician.

⁹From the time of the first VL > 1000 copies/mL result on the DTG-based regimen to the application date for HIVDR testing.

Table 2.	Summary of Genotypic HIVDR Test Results	HIVDR Test Results					
Client No.	VL Result at Time of HIVDR Testing Application	Details INSTI Resistance Mutations	Summary DTG Resistance [12]	Details RT Resistance Mutations	Summary NRTI Resistance	Summary NNRTI Re- sistance	Summary Pl Resistance
~	46 100	Major: R263K; Acces- sory: M50I	Intermediate resistance; mutation score 30	NRTI: M184V/I; NNRTI: H221I	TDF, AZT susceptible; high-level resistance 3TC; low-level resist- ance ABC	Potential low- resistance EFV, low-resistance NVP	ATV, LPV, DRV susceptible
7	29 990	Major: R263K; Accessory: E157Q	Intermediate resistance; mutation score 40	NRTI: M41L, D67N, T69D, K70KN, V75M, M184V, T215F; NNRTI: K103N, V108I, G190A	Intermediate-resistance TDF; high-level resistance ABC, AZT, 3TC	High-level resistance NVP and EFV	ATV, LPV, DRV susceptible
m	000 698	Major: E138K, S147G, R263K, Accessory: A49G, Q95K, E157Q	High-level resist- ance; mutation score 60	NRTI: D67 deletion, T69G, K70R, L74I, M184V, T215V, K219E; NNRTI: A98G, V108I, G190S	Intermediate-resistance TDF; high-level resistance ABC, AZT, 3TC	High-level resistance NVP and EFV	ATV, LPV, DRV susceptible
4	53 943	Major: R263K; Acces- sory: E1570	Intermediate resistance; mutation score 40	NRTI: M41L; D67N, T69G, K70R, M184V/I, T215Y, K219Q/E. NNRTI: V108I/V	Intermediate-resistance TDF; high-level resistance ABC, AZT, 3TC	High-level resistance NVP; intermediate- resistance EFV	ATV, LPV, DRV susceptible
IJ	444 921	Major: S147G; Acces- sory: H51Y	Low-level resistance; mutation score 20	NRTI: K70Q; M184V; NNRTI: Y188HL; P225H	High-level resistance 3TC; low-level resistance TDF; intermediate- resistance ABC; susceptible AZT	High-level resistance NVP and EFV	ATV, LPV, DRV susceptible
Q	4 424 530	Major: R263K; Acces- sory: none	Intermediate resistance; mutation score 30	NRTI: M184V; NNRTI: none	High-level resistance 3TC; low-level resistance ABC; susceptible TDF, AZT	Susceptible EFV, NVP	ATV, LPV, DRV susceptible
	248 541	Major: R263K; Acces- sory: none	Intermediate resistance; mutation score 30	NRTI: D67N, M184V, T215F K219E; NNRTI: A98G, E138A, Y181V	Low-level resistance TDF; intermediate-level resistance ABC; high-level resistance AZT, 3TC	High-level resistance NVP; intermediate resistance EFV	ATV, LPV, DRV susceptible
ω	88 500	Major: R263K; Acces- sory: none	Intermediate resistance; mutation score 30	Not determined	Not determined	Not determined	Not determined
All sample	All samples exhibited HIV subtype C.						

Abbreviations: 3TC, lamivudine; ABC, abacavir, ART, antiretroviral therapy; ATV, atazanavir, AZT, zidovudine; DRV, darunavir; DTG, dolutegravir, EFV, efavirenz; HIVDR, HIV drug resistance; IAC, intensive adherence counseling; INSTI, integrase strand transfer inhibitor; Integrase strand transfer inhibitor; Integrase inhibitor; NL, not applicable; NNRTI, non-nucleoside reverse inhibitor; NVP, nevirapine; PI, protease inhibitor; I, ritonavir; TUF, tenofovir; VL, viral load.

have had undetected HIV minority variants with PI resistance mutations (archived resistance).

HIVDR testing can prevent unnecessary switching to alternative regimens in patients who are not adherent and have no significant HIVDR [17]. It also allows identification of patients with dolutegravir resistance who can benefit from a switch of regimen or from doubling the dolutegravir dose, which has been associated with successful outcomes in ARTexperienced patients who harbored HIV with integrase inhibitor resistance mutations due to previous treatment with raltegravir or elvitegravir [18]. Study of the outcomes of such patients on their modified ART regimens is needed to gain better understanding of dolutegravir resistance mutations and clinical outcomes in African PWH. Local HIVDR capacity is also essential for regular surveillance of dolutegravir resistance development in the Malawi national program. Due to the required high-level expertise and the high costs, HIVDR testing capacity is currently very limited in Sub-Saharan Africa, and integrase sequencing is not yet available for individual clinical care within Malawi. The number of Malawi PWH who require HIVDR testing for their individual management, as current national guidelines require, is therefore much greater than the available laboratory capacity, and it is uncertain if genotyping for all clients failing dolutegravirbased regimens is feasible within the public health approach to ART. More research is needed to establish the exact role of genotyping in settings such as ours. There are many areas of uncertainty about management of patients with persistent viremia on dolutegravir-based regimens in settings where HIVDR capacity is limited or unavailable [19]. Knowledge gaps include the optimal duration of adherence support measures before consideration of HIVDR testing, ART switch decisions in the absence of HIVDR test results, and the best next-line/alternative regimens.

Six of the 33 samples failed to amplify before sequencing, which may be because DBS samples were used for transport to South Africa due to logistical and cost considerations. Another limitation of our survey is that the proportion of dolutegravir resistance we observed may not be extrapolated to the population of Malawians with virological failure on dolutegravir, because the number of individuals who underwent dolutegravir HIVDR testing was very small and likely overrepresented clients from large health facilities.

CONCLUSIONS

We have presented 8 cases with dolutegravir resistance from the Malawi HIV treatment program, where risk factors for dolutegravir resistance are prevalent. These findings advocate for the establishment of adequate HIVDR testing capacity in Malawi to support individual clinical management and regular dolutegravir HIVDR surveillance nationally.

Acknowledgments

J. J. van Oosterhout, C. Chipungu, L. Nkhoma, H. Kanise, M. C. Hosseinipour, J. B. Sagno, K. Simon, S. Phiri, K. Steegen, C. Cox, A. Jahn, and T. Heller collected, reviewed, and interpreted the data. K. Steegen oversaw the laboratory testing. J. J. van Oosterhout wrote the first draft of the manuscript. All authors critically reviewed and agreed with the final draft of the manuscript. We thank the clinicians who referred their clients in the Malawi HIV program for HIVDR testing, the results of which formed the basis of our findings.

Financial support. HIVDR testing in the national HIV program is supported by the President's Emergency Plan for AIDS Relief, through the United States Agency for International Development and Centers for Disease Control and Prevention.

Potential conflicts of interest. The authors have no conflicts of interest. All authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References

- World Health Organization. Update of Recommendations on First- and Second-Line Antiretroviral Regimens. World Health Organization; 2019. Available at: https://apps.who.int/iris/bitstream/handle/10665/325892/WHO-CDS-HIV-19.15-eng.pdf. Accessed 6 April 2022.
- Inzaule SC, Hamers RL, Doherty M, et al. Curbing the rise of HIV drug resistance in low-income and middle-income countries: the role of dolutegravir-containing regimens. Lancet Infect Dis 2019; 19:e246–52.
- Venter WDF, Moorhouse M, Sokhela S, et al. Dolutegravir plus two different prodrugs of tenofovir to treat HIV. N Engl J Med 2019; 381:803–15.
- Kouanfack C, Mpoudi-Etame M, Bassega OP, et al; NAMSAL ANRS 12313 Study Group. Dolutegravir-based or low-dose efavirenz-based regimen for the treatment of HIV-1. N Engl J Med 2019; 381:816–26.
- Lübke N, Jensen B, Hüttig F, et al. Failure of dolutegravir first-line ART with selection of virus carrying R263K and G118R. N Engl J Med 2019; 381:887–9.
- Rhee SY, Grant PM, Tzou PL, et al. A systematic review of the genetic mechanisms of dolutegravir resistance. J Antimicrob Chemother 2019; 74:3135–49.
- Malawi Ministry of Health. Clinical Management of HIV in Children and Adults: Malawi Integrated Guidelines. Malawi Ministry of Health; 2018.
- Malawi Ministry of Health. Malawi Antiretroviral Treatment Program Quarterly Report. Results up to March 31, 2021. Malawi Ministry of Health; 2021.
- Heller T, Ganesh P, Gumulira J, et al. Successful establishment of third-line antiretroviral therapy in Malawi: lessons learned. Public Health Action 2019; 9:169–73.
- Zhou Z, Wagar N, DeVos JR, et al. Optimization of a low cost and broadly sensitive genotyping assay for HIV-1 drug resistance surveillance and monitoring in resource-limited settings. PLoS One 2011; 6:e28184.
- Van Laethem K, Schrooten Y, Covens K, et al. A genotypic assay for the amplification and sequencing of integrase from diverse HIV-1 group M subtypes. J Virol Methods 2008; 153:176–81.
- Tang MW, Liu TF, Shafer RW. The HIVdb system for HIV-1 genotypic resistance interpretation. Intervirology 2012; 55:98–101.
- Temfack E, Jahn A, Kalua T, et al. Prospective enhanced monitoring of dolutegravir-based first line in Malawi (Abstract 35-O-03). Paper presented at: Conference on Retroviruses and Opportunistic Infections; 8–11 March 2020; Boston, MA.
- Paton NI, Musaazi J, Kityo C, et al. NADIA trial team. Dolutegravir or darunavir in combination with zidovudine or tenofovir to treat HIV. N Engl J Med 2021; 385:330–41.
- Neuhann F, de Forest A, Heger E, et al. Pretreatment resistance mutations and treatment outcomes in adults living with HIV-1: a cohort study in urban Malawi. AIDS Res Ther **2020**; 17:22.
- Siedner MJ, Moorhouse MA, Simmons B, et al. Reduced efficacy of HIV-1 integrase inhibitors in patients with drug resistance mutations in reverse transcriptase. Nat Commun 2020; 11:5922.
- Ndahimana Jd, Riedel DJ, Muhayimpundu R, et al. HIV drug resistance mutations among patients failing second-line antiretroviral therapy in Rwanda. Antivir Ther 2016; 21:253–9.
- Castagna A, Maggiolo F, Penco G, et al; VIKING-3 Study Group. Dolutegravir in antiretroviral-experienced patients with raltegravir- and/or elvitegravir-resistant HIV-1: 24-week results of the phase III VIKING-3 study. J Infect Dis 2014; 210:354–62.
- da Silva J, Pals S, Chang J, et al. Monitoring emerging HIV drug resistance in Sub-Saharan Africa in the era of dolutegravir. J Infect Dis 2022; 225:364–6.